SPECIMEN PREPARATION

[Nasopharyngeal swab specimen] specimen preparation for immediate testing

- 1 Insert a sterile nasopharyngeal swab into the nostril of the patient.
- **2** Rotate the swab over the posterior nasopharynx surface 3~4 times.
- 3 Insert the swab into an extraction buffer tube. While squeezing the tube, stir the swab more than 5 times.
- Remove the swab while squeezing the sides of the tube. Press the nozzle cap tightly onto the tube.



NowCheck S COVID-19 Ag Test

For in vitro diagnostics use only

Doc. No.: I1901-10E

PRINCIPLE

NowCheck COVID-19 Ag Test is a rapid chromatographic immunoassay for the qualitative detection of specific SARS-CoV-2 antigens present in human nasopharynx. This test is for administration by healthcare workers and labs only, as an aid to early diagnosis of COVID-19 in patients that are suspected to have a SARS-CoV-2 infection. It provides only an initial screening test result. More specific alternative diagnosis methods should be performed in order to obtain the confirmation of SARS-CoV-2 infection.

NowCheck COVID-19 Ag Test has two pre-coated lines, "C" Control line, "T" Test line on the surface of the nitrocellulose membrane. Both the control line and test line in the result window are not visible before applying any specimens. Mouse monoclonal anti-SARS-CoV-2 antibody is coated on the test line region and mouse monoclonal anti-Chicken IgY antibody is coated on the control line region. Mouse monoclonal anti-SARS-CoV-2 antibody conjugated with color particles is used as detectors for SARS-CoV-2 antigen. During the test, SARS-CoV-2 antigen in the specimen interacts with mouse monoclonal anti-SARS-CoV-2 antibody conjugated with color particles, making antigen-antibody color particle complex. This complex migrates on the membrane via capillary action until the test line, where it will be captured by the mouse monoclonal anti-SARS-CoV-2 antibody. A colored test line would be visible in the result window if SARS-CoV-2 antigens are present in the specimen. The intensity of colored test line will vary depending upon the amount of SARS-CoV-2 antigen present in the specimen. If SARS-CoV-2 antigens are not present in the specimen, then no color appears in the test line. The control line is used for procedural control, and should always appear if the test procedure is performed properly and the test reagents of the control line are working.

MATERIALS PROVIDED

	Reagent	25 Tests/Kit
1	Test device	25
2	Extraction buffer tube	25
3	Nozzle cap	25
4	* Nasopharyngeal swab (Cat. No.: RG1901DG)	25
5	Paper stand	1
6	Film	1
7	Instructions for use	1

* Manufacturer and Authorized representative information for sterile swab is as below.

Noble Biosciences, Inc. 13-50, Sinbaek-gil, Jeongnam-myeon, Hwaseong-si, Gyeonggi-do, Korea Authorized Representative S.B. PHARMA GMBH

Bunsenstr, 14 53121 Bonn, Germany

MATERIALS REQUIRED, BUT NOT PROVIDED

1. Time

2.

VTM

3. Micropipette

STORAGE AND STABILITY

- 1. Store the kit at room temperature (2~30°C / 36~86°F).
- 2. Store the kit out of direct sunlight.
- 3. Do not freeze the kit.
- 4. Shelf life is 24 months. Kit materials are stable until the expiration date printed on the outer box.

PRECAUTIONS

- 1. Do not reuse the test kit.
- 2. Do not use the test kit if the pouch is damaged or the seal is broken.
- 3. Do not use the buffer of another lot.
- 4. Use the test device immediately once taken out of the foil pouch.
- 5. Do not smoke, drink, or eat while handling the specimen or kit reagents.
- Handle all specimens with caution as if they contain infectious agents.
 Wear personal protective equipment, such as gloves and lab coats, when
- handling the specimen and kit reagents. Wash hands thoroughly after the tests are done.
- 8. Clean up spills thoroughly using an appropriate disinfectant.
- 9. The used test and all specimens should be discarded as biohazard waste and must be handled according to local regulations.

- 10. Observe established precautions against microbiological hazards throughout testing procedures.
- 11. Do not use the kit if the test result with positive/negative control is abnormal.

COLLECTION AND PREPARATION OF SPECIMEN

- 1. Specimen collection using a nasopharyngeal swab
 - Insert a nasopharyngeal swab into the nostril of the patient, reaching the surface of the posterior nasopharynx.
 - (2) Using gentle rotation, push the swab until resistance is met at the level of the turbinate.
 - ③ Rotate the swab 3~4 times against the nasopharyngeal wall.
 - ④ Remove the swab from the nostril carefully.
- 2. Specimen should be tested as soon as possible after collection.
- Use the collected specimen and extraction buffer immediately. Be careful of contamination.
- If the specimen cannot be tested immediately after collection, viral transport medium (VTM)* could be used instead of extraction buffer.
- * As the sensitivity of this test can be affected by excessive dilution, it is recommended to use 1 ml VTM.
- 5. The specimen storage condition is as follows.

Specimen Storage Condition	5±3°C	20±5°C
Extraction buffer	4 hours	1 hour
Nasopharyngeal swab inoculated in VTM	12 hours	8 hours

INTERPRETATION OF THE RESULT

- 1. A colored band will appear in the top section of the result window to show that the test is working properly. This band is a control line (C).
- A colored band will appear in the lower section of the result window. This band is a test line of SARS-CoV-2 antigen (T).
- Even if the control line is faint, or the test line isn't uniform, the test should be considered to be performed properly and the test result should be interpreted as positive.
- * The presence of any line no matter how faint the result is considered positive.
- * Positive results should be considered in conjunction with the clinical history and other data available.

SPECIMEN COLLECTION AND TRANSPORT

Commercially available transport medium

Virus Transport Modium (V/TM)	Recommended Storage Condition		
	5±3°C	20±5°C	
UTM [™] (COPAN Diagnostics Inc.)	12 hours	8 hours	
Universal Viral Transport (BD™)	12 hours	8 hours	
FA TRANSPORT MEDIUM (FA Inc.)	12 hours	8 hours	



Allow the VTM containing the specimen to reach room temperature (15-30°C) prior to testing. Refrigerated specimen may fail to move through the device, causing erroneous or invalid results.

PERFORMANCE CHARACTERISTIC

1. Clinical evaluation

Performance characteristics for the NowCheck COVID-19 Ag Test for rapid detection of SARS-CoV-2 antigen was established in a prospective study at a Community Testing Clinic in Brazil during the 2020 SARS-CoV-2 pandemic. A total of 400 prospective specimens were tested using the NowCheck COVID-19 Ag test. These specimens consisted of nasopharyngeal swabs from symptomatic patients. The performance of the NowCheck COVID-19 Ag Test was compared to a Reference RT-PCR method (Lab-developed assay based on the US CDC protocol, which targets two regions(N1 and N2) of the nucleocapsid(N) gene of SARS-CoV-2, using nasopharyngeal swabs). This clinical evaluation was conducted by FIND (Geneva, Switzerland) and its partners, over the period of July – August 2020.

The NowCheck COVID-19 Ag Test showed 89.2% of sensitivity (92.2% sensitivity when the days from symptom onset were \leq 7 days) and 97.3% of specificity, compared to diagnostic RT-PCR assay.

Specimens from symptomatic patients (N=400)		Real-time PCR		
		Positive	Negative	Total
	Positive	91	8	99
NowCheck COVID-19 Ag Test	Negative	11	290	301
	Total	102	298	400
Clinical Sensitivity = 89.2% (95%CI, 81.7% - 93.9%) 92.2% (95%CI, 84.8% - 96.2%) (Days ≤ 7) 91.4% (95%CI, 83.9% - 95.6%) (Ct ≤ 33) Clinical Specificity = 97.3% (95%CI, 94.8% - 98.6%)				

ANALYTICAL PERFORMANCE

 Limit of Detection (LoD): The study used "SARS-CoV-2 (2019-nCOV) NCCP 43326/2020 /Korea" strain. The titer of the cultured virus was confirmed by PCR. The inactivated virus was spiked into the negative nasopharyngeal swab. The LoD is 3.12 X 10²² TCID₅₀/ml.

SARS-CoV-2 strain tested	Virus stock titer	Specimen type	LoD (Spiking titer)	Final working titer	Call rates of 20 replicates near cut-off
NCCP 43326/ 2020 /Korea	1 X 10 ^{6.2} TCID ₅₀ /ml	Direct nasopharyngeal swab	3.12 X 10 ^{2.2} TCID ₅₀ /ml	6.24 X 10 ^{1.2} TCID ₅₀ /ml	100% (20/20)

2. Cross-Reactivity: SARS-CoV showed cross-reactivity, while the others did not show any cross-reactivity at high concentration.

Microbial Organisms	Test titer/value	Result
MERS-Coronavirus	4 X 104 TCID50/ml	No cross-reacitivity
Human Coronavirus 229E	1 X 10 ^{4.5} TCID ₅₀ /ml	No cross-reacitivity
Human Coronavirus OC43	1 X 10 ⁵ TCID ₅₀ /ml	No cross-reacitivity
Human Coronavirus NL63	1 X 104 TCID ₅₀ /ml	No cross-reacitivity
Influenza A	3 X 10 ⁵ TCID ₅₀ /ml	No cross-reacitivity
Influenza B	3 X 10 ⁵ TCID ₅₀ /ml	No cross-reacitivity
RSV A	3 X 10 ⁵ TCID ₅₀ /ml	No cross-reacitivity
RSV B	3 X 10 ⁵ TCID ₅₀ /ml	No cross-reacitivity
Human Metapneumovirus (hMPV)	1 X 10 ⁵ TCID ₅₀ /ml	No cross-reacitivity
Parainfluenza virus	1 X 10 ⁵ TCID ₅₀ /ml	No cross-reacitivity
Rhinovirus	1 X 10 ⁵ TCID ₅₀ /ml	No cross-reacitivity
Enterovirus	1 X 104 TCID ₅₀ /ml	No cross-reacitivity
Adenovirus	3 X 10 ⁵ TCID ₅₀ /ml	No cross-reacitivity
Human immunodeficiency virus lysate	10 µg/ml	No cross-reacitivity
Mycobacterium tuberculosis	5 X 10 ⁴ cells/ml	No cross-reacitivity
Haemophilus influenzae	5 X 10 ⁴ cells/ml	No cross-reacitivity
Mycoplasma pneumoniae	5 X 10 ⁴ cells/ml	No cross-reacitivity
Streptococcus pneumonia	5 X 10 ⁴ cells/ml	No cross-reacitivity
Streptococcus pyrogens	5 X 10 ⁴ cells/ml	No cross-reacitivity
Legionella pneumophila	5 X 10 ⁴ cells/ml	No cross-reacitivity
Candida albicans	5 X 10 ⁴ cells/ml	No cross-reacitivity
Bordetella pertussis	5 X 10 ⁴ cells/ml	No cross-reacitivity
Moraxella catarrhalis	5×10^4 cells/ml	No cross-reacitivity
Pseudomonas aeruginosa	5×10^4 cells/ml	No cross-reacitivity
Staphylococcus epidermidis	5 X 10 ⁴ cells/ml	No cross-reacitivity
Streptococcus salivarius	5 X 10 ⁴ cells/ml	No cross-reacitivity
Chlamydia pneumoniae	1 X 10 ⁵ cells/ml	No cross-reacitivity
Pooled human nasal wash	N/A	No cross-reacitivity

*Human coronavirus HKU1 has not been tested. The % identity of the nucleocapsid protein sequence between HKU1 and SARS-CoV-2 is below 35%.

3. Endogenous/Exogenous Interference Substances Studies: The substances having potential interference are listed in the table below. There was no any interfering activity at high concentration.

Category	Interfering Substances	Test Concentration
	Zanamivir (Influenza)	5 mg/ml
	Oseltamivir (Influenza)	0.039 mg/dL
	Artemether-lumefantrine (Malaria)	50 µM
Polovant modicinos	Doxycycline hyclate (Malaria)	70 µM
Relevant medicines	Quinine (Malaria)	150 µM
	Lamivudine (Retroviral medication)	1.05 mg/dL
	Ribavirin (HCV)	1 mg/ml
	Daclatasvir (HCV)	1 mg/ml
A	Acetaminophen	1030 µM
Anti-Inflammatory medication	Acetylsalicylic acid	167 µM
medication	Ibuprofen	1060 µM
	Mupirocin	3 µM
Antibiotics	Tobramycin	70.6 µM
AITUDIOUCS	Erythromycin (antibiotic)	188 µM
	Ciprofloxacin (antibiotic)	36.2 µM
	Neo-Synephrine (Phenylephrine)	10% (v/v)
	Afrin Nasal Spray (Oxymetazoline)	10% (v/v)
Nasal sprays or drops	Saline Nasal Spray	10% (v/v)
	Rhinocort (Nasal corticosteroids - Budesonide)	10% (v/v)

	Homeopathic Zicam Allergy Relief Nasal Gel	5% (v/v)
Homeopathic allergy relief medicine	Sodium Cromoglycate	20 mg/ml
	Olopatadine Hydrochloride	10 mg/ml
Oral anaesthetic	Anbesol (Benzocaine 20%)	5% (v/v)
Threat lazanges	Strepsils (flurbiprofen 8.75 mg)	5% (w/v, 50 mg/ml)
Throat lozenges	Thoat candy (mint)	5% (w/v, 50 mg/ml)
Others	Mucin: bovine submaxillary gland, type I-S	100 µg/ml
Others	Biotin	14.3 µM
Autoimmuno disease	Human anti-mouse antibody	802 ng/ml
Autoimmune uisease	Rheumatoid factor	3,480 IU/mL
Corum protoin	Whole Blood (human), EDTA anticoagulated	10% (w/w)
Serum protein	Human serum albumin	60 mg/ml

- High-dose Hook Effect: The highest concentration of heat and chemical inactivated SARS-CoV-2 stock available (TCID₅₀ of 1 X 10^{6.2} per ml) was tested. There was no hook effect detected.
- SARS-CoV-2 was inactivated (non-CPE) by the extraction buffer of NowCheck COVID-19 Ag Test in 2 minutes.

Туре	Virus Spiking	Cytopathic Effect	Interpretation
Extraction buffer	0	No CPE	Virus inactivated
Cell culture media	0	CPE	Positive control

 Matrix Equivalency: The matrix and VTM does not affect the detection of COVID-19 Ag in contrived specimen between direct nasopharyngeal swab, nasopharyngeal swab in VTM, direct nasal swab, nasal swab in VTM. (comparator : direct nasopharyngeal swab sample)

LIMITATIONS OF THE TEST

- 1. The test procedure, precautions and interpretation of results for this test must be followed strictly when testing.
- 2. The test should be used for the detection of SARS-CoV-2 antigen in human nasopharyngeal swab.
- 3. Neither the quantitative value nor the rate of SARS-CoV-2 antigen concentration can be determined by this qualitative test.
- Failure to follow the test procedure and interpretation of test results may adversely affect test performance and/or produce invalid results.
- 5. For more accuracy of immune status, additional follow-up testing using other laboratory methods is recommended.
- 6. The test result must always be evaluated with other data available to the physician.
- A negative result may occur if the concentration of antigen in a specimen is below the detection limit of the test or if the specimen is collected or transported improperly. Therefore, a negative test result does not eliminate the possibility of SARS-CoV-2 infection, and should be confirmed by viral culture, a molecular assay, or ELISA.
- 8. Positive test results do not rule out co-infections with other pathogens.
- 9. Negative test results are not intended to rule in other coronavirus infection except the SARS-CoV-1.
- 10. Children tend to shed virus for longer periods than adults, which may result in differences in sensitivity between adults and children.
- 11. When using VTM, sensitivity can be reduced due to excessive dilution.

EXTERNAL QUALITY CONTROL

- Positive and negative controls are optional contents (NowCheck COVID-19 Ag Control(Cat No. RG1901CD)) and these controls can be provided as a means on additional quality control to demonstrate a positive or negative reaction.
- 2. Quality controls should be treated and tested the same as patient specimens.
- 3. It is recommended that positive and negative controls be run: - once for each new lot.
 - once for each untrained operator.
 - as required by test procedures in this instructions and in accordance with local, state, and federal regulations or accreditation requirements.

BIBLIOGRAPHY OF SUGGESTED READING

- 1. Clinical management of severe acute respiratory infection when novel coronavirus(nCoV) infection is suspected. Interim guidance. WHO.2020
- Diagnostic detection of Wuhan coronavirus 2019 by real-time RT-PCR.2020
 Diagnosis and treatment of pneumonia caused by new coronavirus (trial
- Diagnosis and treatment of pneumonia caused by new coronavirus (tria version 4) National Health Commission. 2020

SYMBOL

Symbol	Description
	Manufacturer
Ĩ	Consult instructions for use
REF	Reference number
~~	Date of manufacture To indicate the date of analyzer manufacture
\$	Note
T	Contains sufficient for <n> tests</n>
\otimes	Do not re-use
$\mathbf{\Sigma}$	Use by
\triangle	Caution! Indicates a situation, which if not avoided could result
Ť	Indicates that you should keep the product dry
Ţ	Indicates that the product is fragile and to handle it with care
LOT	Batch code To indicate the lot number
X	Indicates to discard it separately from other household waste
紊	Keep away from sunlight
	Do not use if package is damaged
CE	Fulfill the requirements of Directive 98/79/EC on <i>in vitro</i> diagnostic medical devices
EC REP	Indicates the Authorized Representative in the European Community

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